

the equilibrium concentration of $\text{CH}_2=\text{CHCH}_2\text{SR}$ was of about the same magnitude as the estimated uncertainty in measuring the concentrations;⁹ furthermore, steric effects should be smaller with SR than with SOR, SO_2R , or OR. In the series $n\text{-C}_n\text{H}_{2n+1} > i\text{-Pr} \approx t\text{-Bu}$ it is again not clear that the observed differences are significant, and it is a series in which steric crowding of the CH_2 group in

$\text{XCH}_2\text{CH}=\text{CHY}$ should be less than with $\text{O}i\text{-Bu}$, $\text{SO}_2i\text{-Bu}$, etc. groups. Steric hindrance can decrease resonance interactions with the double bond, as in the first part of the series 2-naphthyl > 1-naphthyl \approx 9-anthryl, but when there is too much hindrance, this resonance effect is counteracted, presumably by crowding the CH_2 group, as in the last part of the series.

Notes

The Periodination Reaction: Fast One-Step Synthesis of C_6I_6 from C_6H_6

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In an attempt to prepare the unknown compound, periodyl benzene (PhIO_3), benzene was added dropwise over a period of around 15 min to a 1.0 M solution of H_5IO_6 in concentrated H_2SO_4 in an open beaker at 0–5 °C, whereupon the colorless solution turns green,¹ then red, and finally light yellow, as a yellow-tan precipitate gradually forms, which, after recrystallization from Me_2SO , is insoluble in all common solvents except Me_2SO and MeCN : mp ~ 260 °C with decomposition, giving off I_2 ; elemental analysis, 8.5% C and 91.5% I; $M_r \approx 800$ by freezing point depression of camphor; mass spectrum parent peak at 834 (C_6I_6^+) and $M - 1$ at 707 (C_6I_5^+); proton NMR, no resonance absorption; burns with an aromatic sooty flame along with dense purple fumes of I_2 , from all of which evidence one would rightly conclude that the compound prepared here is C_6I_6 ,² and, on the basis of the quantity of benzene used, the yield is 48% periodobenzene.

Registry No. H_5IO_6 , 10450-60-9; C_6I_6 , 608-74-2; PhIO_3 , 82891-66-5; benzene, 71-43-2.

(1) The green intermediate first formed (presumably PhIO_3), and the red one, should be further investigated, as well as the generality of the periodination of aromatics. The authors invite any investigator interested in this unusual reaction, which might be of use in deuterating aromatic compounds, to pursue this research.

(2) The stoichiometric equation used to calculate yield is $2\text{C}_6\text{H}_6 + 3\text{IO}_4^- + 9\text{I}^- + 12\text{H}_3\text{O}^+ \rightarrow 2\text{C}_6\text{I}_6 + 24\text{H}_2\text{O}$, the I^- indicating that some of the benzene is oxidized, presumably to CO_2 .

Stereoselective Synthesis of (23*S*,25*R*)-23,25,26-Trihydroxyvitamin D₃ and (23*S*,25*R*)-25-Hydroxyvitamin D₃ 26,23-Lactol, Presumed Vitamin D₃ Metabolites

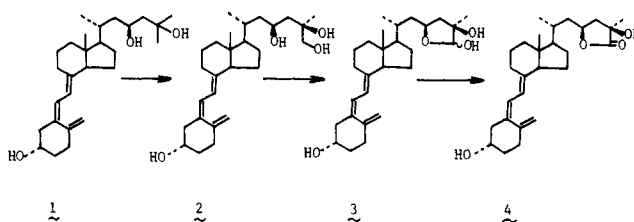
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Calcidiol lactone, 25-hydroxyvitamin D₃ 26,23-lactone (4),¹ is a unique metabolite of vitamin D₃ which exhibits

Scheme I. Presumed Metabolic Pathway of (23*S*)-23,25-Dihydroxyvitamin D₃ (1) to Calcidiol Lactone (4)



a weak activity in intestinal calcium transport and bone calcium mobilization but shows the most potent activity² toward vitamin D binding protein in blood plasma of all known vitamin D metabolites. These characteristics have suggested that the metabolite may have an important role in other aspects of vitamin D action. As one of our projects on the stereoselective synthesis of vitamin D metabolites using chiral templates,³ we have synthesized (23*R*,25*S*)-⁴ and (23*S*,25*R*)-calcidiol lactones⁵ stereoselectively and for the first time determined the stereochemistry of the natural metabolite⁵ to be *S* at C-23 and *R* at C-25. Recently a new metabolite, (23*S*)-23,25-dihydroxyvitamin D₃ (1),⁶ has been isolated and has been shown to be a biosynthetic precursor of calcidiol lactone (4).⁷ It can be assumed that biological transformation of 23,25-dihydroxyvitamin D₃ (1) to the lactone (4) may proceed via 23,25,26-trihydroxyvitamin D₃ (2) through 25-hydroxyvitamin D₃ 26,23-lactol (3; Scheme I) and that these postulated biosynthetic intermediates have the same stereochemical configuration at C-23 and C-25 as those of calcidiol lactone (4). So we planned the stereoselective synthesis of these two presumed vitamin D₃ metabolites.

In this paper we report the stereoselective synthesis of (23*S*,25*R*)-23,25,26-trihydroxyvitamin D₃ (2) and (23*S*,25*R*)-25-hydroxyvitamin D₃ 26,23-lactol (3). Both compounds have been demonstrated to be converted to

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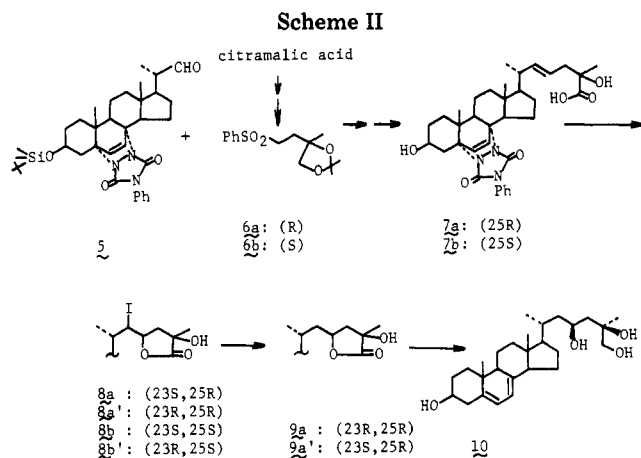
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calcidiol lactone (4) by *in vitro* incubation with chick kidney homogenates, as expected.^{8,9}

The strategy for the synthesis of the two compounds (2 and 3) is essentially the same as that for calcidiol lactone;^{4,5} an *R* configuration at C-25 is introduced by utilizing the chirality of (*R*)-(-)-citramalic acid, and the *S* configuration at C-23 is induced by the stereoselective iodolactonization of the Δ^{22} -26-carboxylic acid 7a. The (25*R*)-carboxylic acid 7a^{4,5} was synthesized by starting with C(22)-steroid aldehyde 5 and optically pure (*R*)-sulfone 6a (Scheme II), which was readily obtained from (*R*)-citramalic acid in eight steps in 67% overall yield.^{4,5}

Iodolactonization of the unsaturated carboxylic acid 7 was studied in some detail to induce the desired chirality at C-23. It has been reported by Barton et al.¹⁰ that the iodoacetoxylation of steroids with a 22(23) double bond proceeds in a regio- and stereoselective manner to yield the iodoacetate in which the bulky iodine is introduced at C-22 from the sterically less hindered side of the molecule. In accord with their results, the iodolactonization (I_2 , CH_3CN) of Δ^{22} -steroidal carboxylic acid 11 has been reported¹¹ to yield exclusively the (23*S*)-iodolactone 12 (Scheme III). It was also found, in our stereoselective synthesis of (23*R*,25*S*)-calcidiol lactone,⁴ that the iodolactonization (I_2 , CH_3CN) of (25*S*)-carboxylic acid 7b gave the (23*S*)-iodolactone 8b in 90% stereoselectivity, in accord with the precedents. In the extensive studies on the stereoselective iodolactonization of acyclic γ,δ -unsaturated carboxylic acids,¹² it has been reported that the stereose-

Table I. Iodolactonization of Δ^{22} -26-Carboxylic Acid (7a,b)

entry	sub- strate	conditions	product distribution, %			
			8a	8a'	8b	8b'
1	7a	I_2 , CH_3CN	57	43	90	10
	7b					
2	7a	I_2 , Et_2O -THF, aqueous $NaHCO_3$	43	57	88	12
	7b					
3	7a	I_2 , CH_3CN , pyridine	27	73	57	43
	7b					
4	7a	I_2 , CH_2Cl_2 , pyridine	20	80	55	45
	7b					

lectivity of the reaction depends on the reaction conditions, and under acidic conditions (I_2 , CH_3CN) the thermodynamically more stable isomers are produced selectively while under basic conditions (I_2 , Et_2O -THF, aqueous $NaHCO_3$) thermodynamically less stable isomers, kinetic products, are formed predominantly. We examined the iodolactonization of the two epimeric carboxylic acids 7a and 7b under thermodynamic and kinetic conditions. As Table I shows, the stereoselectivity of the reaction depends on both the stereochemistry at C-25 of the carboxylic acid 7 and the reaction conditions. Although the 25*S* isomer 7b gave the (23*S*)-iodolactone 8b¹³ in high selectivity under acidic conditions as described above, the 25*R* isomer 7a yielded both (23*S*)- and (23*R*)-iodolactones 8a and 8a' in a comparable ratio under the same conditions (entry 1). However, in an attempted iodolactonization of 7a and 7b under the kinetic conditions reported by Chamberlin et al.^{12b} (entry 2), no appreciable change was observed in the ratio of the products. A remarkable change in the stereoselectivity was observed when pyridine was added to the reaction mixture as the base (entry 3 and 4), and in the case of (25*R*)-carboxylic acid 7a the selectivity was reversed, giving rise to the desired iodolactone 8a' in 80% selectivity. It is likely that the function of pyridine in the stereochemical consequence of the reaction may be similar to that of $NaHCO_3$, because it was also found that in a well-studied system,¹² iodolactonization of 3-methyl-4-pentenoic acid, pyridine worked similarly to $NaHCO_3$, yielding the kinetic product predominantly.¹⁴ Although it is not clearly understood why the addition of $NaHCO_3$ was not effective in our system in altering the stereochemical course of the reaction, pyridine is shown to be a useful reagent for the purpose.

The mixture of the (23*R*,25*R*)- and (23*S*,25*R*)-iodolactones 8a' and 8a from the entry 4 experiment was subjected to reduction with *n*- Bu_3SnH without separation. After purification on a silica gel column, the major (23*S*,25*R*)-lactone 9a' was obtained in pure form in 56% overall yield (from 7a). Reduction of the lactone 9a' with $LiAlH_4$ was accompanied by the deprotection of the 5,7-diene group to afford the desired provitamin D (10) in 90% yield. The provitamin D (10) was transformed into the corresponding vitamin D (2) by UV irradiation followed by thermal isomerization as usual.

The synthesis of (23*S*,25*R*)-25-hydroxyvitamin D₃ 26,23-lactol (3) was performed in one step from

(8) We have noticed the presence of a new metabolite which migrated to the same retention volume as that of (23*S*,25*R*)-23,25,26-trihydroxyvitamin D₃ in the incubates of chick kidney homogenate with 25-hydroxyvitamin D₃. Detailed studies on the metabolite are currently progressing.

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(23*S*,25*R*)-caldiol lactone (4)⁵ by reduction with diisobutyl aluminium hydride (DIBAL) (toluene, -70 °C) in high yield (93%). It is interesting to note that only one anomer at C-26 was obtained as verified by the ¹H NMR spectrum (CDCl₃): δ 1.34 (3 H, s, H-27) 3.8-4.20 (2 H, m, H-3 and H-23), 4.83 (2 H, br s, H-19 and H-26), 5.07 (1 H, br s, H-19), 6.14 (2 H, AB q, *J* = 11 Hz, H-6 and H-7).

Experimental Section

The melting point was determined with a Yanaco micro melting point apparatus and was not corrected. ¹H NMR spectra were obtained with a Varian XL-100 instrument. Chemical shifts are reported in parts per million relative to tetramethylsilane. Mass spectra were obtained with a JEOL JMS-D300 spectrometer. Infrared spectra were obtained with a JASCO A-302 spectrometer. UV spectra were recorded on a Union Giken SM 401 spectrometer.

Iodolactonization of 4-Phenyl-1,2,4-triazoline-3,5-dione Adducts of (25*R*)- and (25*S*)-3β,25-Dihydroxy-5,7,22-cholestatrien-26-oic Acids (7a,b). **Method A.** Iodine (25 mg, 9.8 × 10⁻² mmol) was added to a solution of carboxylic acid 7 (20 mg, 3.3 × 10⁻² mmol) in CH₃CN (1 mL) at 0 °C, and the solution was stirred at that temperature for 5 h. After addition of aqueous Na₂S₂O₃, the mixture was extracted with CHCl₃, washed with brine, dried over Na₂SO₄, and evaporated. The product ratio was analyzed by HPLC [column, μ-Porasil; solvent, 2-propanol-hexane (15:85) for the analysis of the products from 7b and ethyl acetate-hexane (7:3) for those from 7a].

Method B. Carboxylic acid 7 (20 mg) was dissolved in Et₂O-THF (1:1 2 mL) and combined with aqueous saturated NaHCO₃ (2 mL). The solution was stirred for 30 min at room temperature and cooled to 0 °C, and then iodine (25 mg) was added. After 5 h, the reaction mixture was worked up as above, and the products were analyzed by HPLC.

Method C. A solution of carboxylic acid 7 (20 mg) and pyridine (20 μL, 2.5 × 10⁻¹ mmol) in CH₃CN or CH₂Cl₂ (1 mL) was stirred at room temperature for 30 min and cooled to 0 °C, and then iodine (25 mg) was added. The mixture was stirred at that temperature for 5 h and then worked up as above. The products were analyzed by HPLC.

The results of the iodolactonization are shown in Table I.

4-Phenyl-1,2,4-triazoline-3,5-dione Adducts of (23*S*,25*R*)- and (23*R*,25*R*)-3β,25-Dihydroxy-22-iodo-5,7-cholestadiene 26,23-Lactone (8a,a'). A solution of 7a (105 mg, 1.74 × 10⁻¹ mmol) and pyridine (105 μL, 1.3 mmol) in CH₂Cl₂ (6 mL) was stirred at room temperature for 30 min and cooled to 0 °C, and then iodine (133 mg, 5.24 × 10⁻¹ mmol) was added. After 2.5 h, iodine (133 mg) and pyridine (105 μL) were added, and the resultant solution was stirred for a further 7.5 h at 0 °C. Aqueous Na₂S₂O₃ was added, and the mixture was extracted with CHCl₃, washed with brine, dried over Na₂SO₄, and evaporated. The residue was chromatographed on silica gel (6 g) by using hexane-ethyl acetate (2:8) as the eluent to give a 1:4 mixture of 8a and 8a' (95 mg). The mixture was subjected to the next reaction without separation.

4-Phenyl-1,2,4-triazoline-3,5-dione Adducts of (23*R*,25*R*)- and (23*S*,25*R*)-3β,25-Dihydroxy-5,7-cholestadiene 26,23-Lactone (9a,a'). To a solution of the iodolactone (8a and 8a', 1:4; 100 mg, 1.37 × 10⁻¹ mmol) in DME (4 mL) was added *n*-Bu₃SnH (300 μL, 1.14 mmol), and the mixture was stirred at 60 °C for 1.5 h. After evaporation of the solvent, the residue was dissolved in CH₃CN and washed with hexane to remove organic tin compounds, and the CH₃CN was evaporated. The residue was chromatographed on silica gel (10 g) with hexane-ethyl acetate (2:8) as the eluent to yield 9a (16 mg) and 9a' (62 mg) in that order. 9a: MS, *m/e* 428 (M⁺ - triazoline), 395. 9a': MS, *m/e* 428 (M⁺ - triazoline), 395; IR (CHCl₃) 1685, 1785, 1770; ¹H NMR (CDCl₃) δ 0.80 (3 H, s, H-18), 0.94 (3 H, s, H-19), 1.44 (3 H, s, H-27), 4.15-4.65 (2 H, m, H-3 and H-23).

(23*S*,25*R*)-5,7-Cholestadiene-3β,23,25,26-tetrol (10). To a suspension of LiAlH₄ (10 mg, 0.26 mmol) in THF (1 mL) was added a solution of 9a' (28 mg, 4.6 × 10⁻² mmol) in THF (1 mL), and the mixture was refluxed for 50 min. After the excess of the reagent was quenched with aqueous THF, the mixture was filtered and washed with THF and CHCl₃-MeOH (2:1), and the combined filtrate and washings were dried over Na₂SO₄ and evaporated. The residue was chromatographed on Sephadex LH-20 (10 g) with

hexane-CHCl₃-MeOH (25:75:2) as the eluent to yield 10: 18 mg; mp 225-228 °C; MS, *m/e* 432 (M⁺), 414, 399, 383; ¹H NMR (Me₂SO-*d*₆) δ 0.58 (3 H, s, H-18), 0.86 (3 H, s, H-19), 1.07 (3 H, s, H-27), 3.6-4.0 (2 H, m, H-3 and H-23), 5.37 and 5.52 (2 H, m, H-6 and H-7); UV (95% EtOH) 272, 282, 293 nm.

(23*S*,25*R*)-23,25,26-Trihydroxyvitamin D₃ (2). A solution of 10 (6 mg) in 95% EtOH (200 mL) was irradiated by a high-pressure mercury lamp (200 W) through a Vycor filter for 5 min under an argon atmosphere, the temperature being maintained below 5 °C. The solvent was evaporated, and the residue was chromatographed on Sephadex LH-20 (25 g) and eluted with hexane-CHCl₃-MeOH (25:75:2.5) to yield previtamin D: 2.6 mg; UV (95% EtOH) 260 nm. The previtamin D was dissolved in 95% EtOH (2 mL), heated for 7 h at 60-63 °C, and then allowed to stand at room temperature for 8 h. After evaporation of the solvent, the residue was chromatographed on Sephadex LH-20 (25 g) and eluted with hexane-CHCl₃-MeOH (25:75:2.5) to give vitamin D 2: 1.9 mg; high-resolution MS, C₂₇H₄₄O₄ requires *m/e* 432.3239, found *m/e* 432.3256; ¹H NMR (CDCl₃) δ 0.56 (3 H, s, H-18), 1.22 (3 H, s, H-27), 3.57 (2 H, AB q, *J* = 11 Hz, H-26), 3.8-4.2 (2 H, m, H-3 and H-23), 4.82 (1 H, br s, H-19), 5.04 (1 H, br s, H-19), 6.13 (2 H, AB q, *J* = 11 Hz, H-6 and -7); UV (95% EtOH) 265 nm.

(23*S*,25*R*)-25-Hydroxyvitamin D₃ 26,23-Lactol (3). A solution of (23*S*,25*R*)-caldiol lactone (4; 1.8 mg, 4.2 × 10⁻³ mmol) in toluene (300 μL) was cooled to -70 °C under argon, diisobutyl aluminium hydride (25% hexane solution, 26.5 μL, 4.7 × 10⁻² mmol) was added, and the mixture was stirred for 2 h at that temperature. The reaction was quenched with cold ethanol (150 μL) at -70 °C, stirred for 20 min at that temperature, and then allowed to warm to room temperature. The mixture was diluted with CHCl₃, washed with 5% HCl and water, dried over Na₂SO₄, and evaporated. The residue was chromatographed on Sephadex LH-20 (10 g) with hexane-CHCl₃-MeOH (35:65:2) as the eluent to give lactol 3: 1.67 mg; MS, *m/e* 430 (M⁺), 412, 394, 379, 356, 342; IR (CHCl₃) 3400 cm⁻¹; UV (95% EtOH) 265 nm.

Registry No. 2, 83198-41-8; 3, 83136-06-5; 4, 77714-47-7; 7a, 81495-58-1; 7b, 80320-87-2; 8a, 80320-88-3; 9a, 78109-10-1; 9a', 78183-86-5; 10, 83136-07-6; (23*S*,25*R*)-23,25,26-trihydroxyprevitamin D₃, 83136-08-7.

Reaction between 2-Amino-2-deoxy-D-glucose Derivatives and Sulfite. 2. Synthesis of 2-(D-*arabino*-Tetrahydroxybutyl)-5-(3,4-dihydroxy-2-sulfobutyl)pyrazine

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In continuation of our studies of the reaction of 2-amino-2-deoxy-D-glucose (1) derivatives in sodium bisulfite solution we recently reported the condensation of 2-amino-2-deoxy-D-glucose oxime with glyoxal-sodium bisulfite.¹ We now report the condensation of 2 mol of 1 in heated sodium bisulfite solution. The principal product is a pyrazine derivative with two polyhydroxyalkyl side chains one of which is sulfonated. The reaction constitutes a new route to sulfonated polyhydroxyalkylpyrazine derivatives.

The formation of the two pyrazine derivatives "fructosazine" (2) and "deoxyfructosazine" (3) from 1 has been reported.^{2,3} Excessive side reactions of 1 in alkaline media resulted, however, in a low yield of 2. Ingles^{4,5} found

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